

1 INDICATIONS AND USAGE

ZERBAXA (ceftolozane and tazobactam) for injection is indicated for the treatment of patients 18 years or older with the following infections caused by designated susceptible microorganisms.

1.1 Complicated Intra-abdominal Infections

ZERBAXA used in combination with metronidazole is indicated for the treatment of complicated intra-abdominal infections (cIAI) caused by the following Gram-negative and Gram-positive microorganisms: *Enterobacter cloacae*, *Escherichia coli*, *Klebsiella oxytoca*, *Klebsiella pneumoniae*, *Proteus mirabilis*, *Pseudomonas aeruginosa*, *Bacteroides fragilis*, *Streptococcus anginosus*, *Streptococcus constellatus*, and *Streptococcus salivarius*.

1.2 Complicated Urinary Tract Infections, Including Pyelonephritis

ZERBAXA is indicated for the treatment of complicated urinary tract infections (cUTI), including pyelonephritis, caused by the following Gram-negative microorganisms: *Escherichia coli*, *Klebsiella pneumoniae*, *Proteus mirabilis*, and *Pseudomonas aeruginosa*.

1.3 Nosocomial Pneumonia, including Ventilator-associated Pneumonia

Treatment of nosocomial pneumonia, including ventilator-associated pneumonia, caused by the following susceptible Gram-negative microorganisms: *Enterobacter cloacae*, *Escherichia coli*, *Haemophilus influenzae*, *Klebsiella oxytoca*, *Klebsiella pneumoniae*, *Proteus mirabilis*, *Pseudomonas aeruginosa*, and *Serratia marcescens*.

1.4 Usage

To reduce the development of drug-resistant bacteria and maintain the effectiveness of ZERBAXA and other antibacterial drugs, ZERBAXA should be used only to treat infections that are proven or strongly suspected to be caused by susceptible bacteria. When culture and susceptibility information are available, they should be considered in selecting or modifying antibacterial therapy. In the absence of such data, local epidemiology and susceptibility patterns may contribute to the empiric selection of therapy.

2 DOSAGE AND ADMINISTRATION

2.1 Recommended Dosage

The recommended dosage regimen of ZERBAXA for injection is 1.5 gram (g) (ceftolozane 1 g and tazobactam 0.5 g) for cIAI and cUTI and 3 g (ceftolozane 2 g and tazobactam 1 g) for nosocomial pneumonia administered every 8 hours by intravenous infusion over 1 hour in patients 18 years or older and with normal renal function or mild renal impairment. The duration of therapy should be guided by the severity and site of infection and the patient's clinical and bacteriological progress as shown in Table 1.

Table 1: Dosage of ZERBAXA by Infection in Patients with Creatinine Clearance (CrCl) Greater than 50 mL/min

Infection	Dose	Frequency	Infusion Time	Duration of Treatment
Complicated Intra-abdominal Infections*	1.5 g	Every 8 Hours	1	4-14 days
Complicated Urinary Tract Infections, including Pyelonephritis	1.5 g	Every 8 Hours	1	7 days
Nosocomial Pneumonia, including Ventilator-associated Pneumonia	3 g	Every 8 Hours	1	8-14 days

* Used in conjunction with metronidazole 500 mg intravenously every 8 hours

2.2 Patients with Renal Impairment

Dose adjustment is required for patients whose creatinine clearance is 50 mL/min or less. Renal dose adjustments are listed in Table 2. For patients with changing renal function, monitor CrCl at least daily and adjust the dosage of ZERBAXA accordingly [see *Use in Specific Populations (7)* and *Clinical Pharmacology (10.3)*].

Table 2: Recommended Dosage Regimens for ZERBAXA in Patients with Renal Impairment

Estimated CrCl (mL/min)*	Complicated Intra-abdominal Infections and Complicated Urinary Tract Infections, including Pyelonephritis [†]	Nosocomial Pneumonia, including Ventilator-associated Pneumonia [†]

30 to 50	750 mg (500 mg and 250 mg) intravenously every 8 hours	1.5 g (1 g and 0.5 g) intravenously every 8 hours
15 to 29	375 mg (250 mg and 125 mg) intravenously every 8 hours	750 mg (500 mg and 250 mg) intravenously every 8 hours
End-stage renal disease (ESRD) on hemodialysis (HD)	A single loading dose of 750 mg (500 mg and 250 mg) followed after 8 hours by a 150 mg (100 mg and 50 mg) maintenance dose administered every 8 hours for the remainder of the treatment period (on hemodialysis days, administer the dose at the earliest possible time following completion of dialysis)	A single loading dose of 2.25 g (1.5 g and 0.75 g) followed after 8 hours by a 450 mg (300 mg and 150 mg) maintenance dose administered every 8 hours for the remainder of the treatment period (on hemodialysis days, administer the dose at the earliest possible time following completion of dialysis)

* CrCl estimated using Cockcroft-Gault formula

† All doses of ZERBAXA are administered over 1 hour.

2.3 Patients with Hepatic impairment

No dose adjustment is necessary in patients with hepatic impairment [see *Clinical Pharmacology (10.3)*].

2.4 Preparation of Solutions

ZERBAXA does not contain a bacteriostatic preservative. Aseptic technique must be followed in preparing the infusion solution.

Preparation of doses:

Constitute each vial of ZERBAXA with 10 mL of sterile water for injection or 0.9% Sodium Chloride for Injection, USP and gently shake to dissolve. The final volume is approximately 11.4 mL per vial. Caution: The constituted solution is not for direct injection.

To prepare the required dose, withdraw the appropriate volume determined from Table 3 from the reconstituted vial(s). Add the withdrawn volume to an infusion bag containing 100 mL of 0.9% Sodium Chloride for Injection, USP or 5% Dextrose Injection, USP.

Table 3: Preparation of Doses

ZERBAXA (ceftolozane and tazobactam) Dose	Volume to Withdraw from Reconstituted Vial(s)
3 g (2 g and 1 g)	Two vials of 11.4 mL each (entire contents from two vials)
2.25 g (1.5 g and 0.75 g)	11.4 mL from one vial (entire contents) and 5.7 mL from a second vial
1.5 g (1 g and 0.5 g)	11.4 mL (entire contents from one vial)
750 mg (500 mg and 250 mg)	5.7 mL
450 mg (300 mg and 150 mg)	3.5 mL
375 mg (250 mg and 125 mg)	2.9 mL
150 mg (100 mg and 50 mg)	1.2 mL

Inspect drug products visually for particulate matter and discoloration prior to use. ZERBAXA infusions range from clear, colorless solutions to solutions that are clear and slightly yellow. Variations in color within this range do not affect the potency of the product.

2.5 Compatibility

Compatibility of ZERBAXA with other drugs has not been established. ZERBAXA should not be mixed with other drugs or physically added to solutions containing other drugs.

2.6 Storage of Constituted Solutions

Upon constitution with sterile water for injection or 0.9% sodium chloride injection, reconstituted ZERBAXA solution may be held for 1 hour prior to transfer and dilution in a suitable infusion bag.

Following dilution of the solution with 0.9% sodium chloride or 5% dextrose, ZERBAXA is stable for 24 hours when stored at room temperature or 7 days when stored under refrigeration at 2 to 8°C (36 to 46°F).

Constituted ZERBAXA solution or diluted ZERBAXA infusion should not be frozen.

3 DOSAGE FORMS AND STRENGTHS

ZERBAXA 1.5 g (ceftolozane and tazobactam) for injection is supplied as a white to yellow sterile powder for reconstitution in single-dose vials; each vial contains ceftolozane 1 g (equivalent to 1.147 g of ceftolozane sulfate) and tazobactam 0.5 g (equivalent to 0.537 g of tazobactam sodium). For doses above 1.5 g, two vials of ZERBAXA are used.

4 CONTRAINDICATIONS

ZERBAXA is contraindicated in patients with:

- Hypersensitivity to the active substances or to any of the inactive excipients;
- Hypersensitivity to any cephalosporin antibacterial agent;
- Severe hypersensitivity (e.g., anaphylactic reaction, severe skin reaction) to any other type of beta-lactam antibacterial agent (e.g., penicillins or carbapenems).

5 WARNINGS AND PRECAUTIONS

5.1 Decreased Efficacy in Patients with Baseline Creatinine Clearance of 30 to ≤ 50 mL/min

In a subgroup analysis of a Phase 3 cIAI trial, clinical cure rates were lower in patients with baseline creatinine clearance (CrCl) of 30 to ≤ 50 mL/min compared to those with CrCl > 50 mL/min (Table 4). The reduction in clinical cure rates was more marked in the ZERBAXA plus metronidazole arm compared to the meropenem arm. A similar trend was also seen in the cUTI trial. Monitor CrCl at least daily in patients with changing renal function and adjust the dosage of ZERBAXA accordingly [see *Dosage and Administration (2.2)*].

Table 4: Clinical Cure Rates in a Phase 3 Trial of cIAI by Baseline Renal Function (MITT Population)

Baseline Renal Function	ZERBAXA plus metronidazole n/N (%)	Meropenem n/N (%)
Normal/mild impairment (CrCl > 50 mL/min)	312/366 (85.2)	355/404 (87.9)
Moderate impairment (CrCl 30 to ≤ 50 mL/min)	11/23 (47.8)	9/13 (69.2)

5.2 Hypersensitivity Reactions

Serious and occasionally fatal hypersensitivity reactions (including anaphylactoid and severe cutaneous adverse reactions) have been reported in patients receiving beta-lactam antibacterial drugs.

Before initiating therapy with ZERBAXA, make careful inquiry about previous hypersensitivity reactions to other cephalosporins, penicillins, carbapenems, or other beta-lactams. If this product is to be given to a patient with a cephalosporin, penicillin, carbapenem, or other beta-lactam allergy, exercise caution because cross sensitivity has been established. If an allergic reaction to ZERBAXA occurs, discontinue the drug and institute appropriate alternative therapy.

5.3 *Clostridium difficile*-associated Diarrhea

Clostridium difficile-associated diarrhea (CDAD) has been reported for nearly all systemic antibacterial agents, including ZERBAXA, and may range in severity from mild diarrhea to fatal colitis. Treatment with antibacterial agents alters the normal flora of the colon and may permit overgrowth of *C. difficile*.

C. difficile produces toxins A and B which contribute to the development of CDAD. CDAD must be considered in all patients who present with diarrhea following antibacterial use. Careful medical history is necessary because CDAD has been reported to occur more than 2 months after the administration of antibacterial agents.

If CDAD is confirmed, discontinue antibacterials not directed against *C. difficile*, if possible. Manage fluid and electrolyte levels as appropriate, supplement protein intake, monitor antibacterial treatment of *C. difficile*, and institute surgical evaluation as clinically indicated.

5.4 Development of Drug-Resistant Bacteria

Prescribing ZERBAXA in the absence of a proven or strongly suspected bacterial infection is unlikely to provide benefit to the patient and risks the development of drug-resistant bacteria.

6 ADVERSE REACTIONS

The following serious reactions are described in greater detail in the Warnings and Precautions section:

- Hypersensitivity reactions [see *Warnings and Precautions* (5.2)]
- *Clostridium difficile*-associated diarrhea [see *Warnings and Precautions* (5.3)]

6.1 Clinical Trial Experience

Because clinical trials are conducted under widely varying conditions, adverse reaction rates observed in the clinical trials of a drug cannot be directly compared to rates in the clinical trials of another drug and also may not reflect rates observed in practice.

Complicated Intra-abdominal Infections and Complicated Urinary Tract Infections, including Pyelonephritis

ZERBAXA was evaluated in Phase 3 comparator-controlled clinical trials of cIAI and cUTI, which included a total of 1015 patients treated with ZERBAXA (1.5 g every 8 hours, adjusted based on renal function where appropriate) and 1032 patients treated with comparator (levofloxacin

750 mg daily in cUTI or meropenem 1 g every 8 hours in cIAI) for up to 14 days. The mean age of treated patients was 48 to 50 years (range 18 to 92 years), across treatment arms and indications. In both indications, about 25% of the subjects were 65 years of age or older. Most patients (75%) enrolled in the cUTI trial were female, and most patients (58%) enrolled in the cIAI trial were male. Most patients (>70%) in both trials were enrolled in Eastern Europe and were White.

The most common adverse reactions (5% or greater in either indication) occurring in patients receiving ZERBAXA were nausea, diarrhea, headache, and pyrexia. Table 5 lists adverse reactions occurring in 1% or greater of patients receiving ZERBAXA in Phase 3 cIAI and cUTI clinical trials.

Table 5: Adverse Reactions Occurring in 1% or Greater of Patients Receiving ZERBAXA in Phase 3 cIAI and cUTI Clinical Trials

Preferred Term	Complicated Intra-abdominal Infections		Complicated Urinary Tract Infections, Including Pyelonephritis	
	ZERBAXA* (N=482) n (%)	Meropenem (N=497) n (%)	ZERBAXA* (N=533) n (%)	Levofloxacin (N=535) n (%)
Nausea	38 (7.9)	29 (5.8)	15 (2.8)	9 (1.7)
Headache	12 (2.5)	9 (1.8)	31 (5.8)	26 (4.9)
Diarrhea	30 (6.2)	25 (5)	10 (1.9)	23 (4.3)
Pyrexia	27 (5.6)	20 (4)	9 (1.7)	5 (0.9)
Constipation	9 (1.9)	6 (1.2)	21 (3.9)	17 (3.2)
Insomnia	17 (3.5)	11 (2.2)	7 (1.3)	14 (2.6)
Vomiting	16 (3.3)	20 (4)	6 (1.1)	6 (1.1)
Hypokalemia	16 (3.3)	10 (2)	4 (0.8)	2 (0.4)
ALT increased	7 (1.5)	5 (1)	9 (1.7)	5 (0.9)
AST increased	5 (1)	3 (0.6)	9 (1.7)	5 (0.9)
Anemia	7 (1.5)	5 (1)	2 (0.4)	5 (0.9)
Thrombocytosis	9 (1.9)	5 (1)	2 (0.4)	2 (0.4)
Abdominal pain	6 (1.2)	2 (0.4)	4 (0.8)	2 (0.4)
Anxiety	9 (1.9)	7 (1.4)	1 (0.2)	4 (0.7)
Dizziness	4 (0.8)	5 (1)	6 (1.1)	1 (0.2)
Hypotension	8 (1.7)	4 (0.8)	2 (0.4)	1 (0.2)
Atrial fibrillation	6 (1.2)	3 (0.6)	1 (0.2)	0

Rash	8 (1.7)	7 (1.4)	5 (0.9)	2 (0.4)
Infusion site reaction	3 (0.6)	6 (1.2)	7 (1.3)	11 (2.1)

*The ZERBAXA for injection dose was 1.5 g intravenously every 8 hours, adjusted to match renal function where appropriate. In the cIAI trials, ZERBAXA was given in conjunction with metronidazole.

Treatment discontinuation due to adverse events occurred in 2.0% (20/1015) of patients receiving ZERBAXA and 1.9% (20/1032) of patients receiving comparator drugs. Renal impairment (including the terms renal impairment, renal failure, and renal failure acute) led to discontinuation of treatment in 5/1015 (0.5%) subjects receiving ZERBAXA and none in the comparator arms.

Increased Mortality

In the cIAI trials (Phase 2 and 3), death occurred in 2.5% (14/564) of patients receiving ZERBAXA and in 1.5% (8/536) of patients receiving meropenem. The causes of death varied and included worsening and/or complications of infection, surgery and underlying conditions.

Less Common Adverse Reactions in Phase 3 cIAI and cUTI Clinical Trials

The following selected adverse reactions were reported in ZERBAXA-treated subjects at a rate of less than 1%:

Cardiac disorders: tachycardia, angina pectoris

Gastrointestinal disorders: gastritis, abdominal distension, dyspepsia, flatulence, ileus paralytic

Infections and infestations: candidiasis including oropharyngeal and vulvovaginal, fungal urinary tract infection

Investigations: increased serum gamma-glutamyl transpeptidase (GGT), increased serum alkaline phosphatase, positive Coombs test

Metabolism and nutrition disorders: hyperglycemia, hypomagnesemia, hypophosphatemia

Nervous system disorders: ischemic stroke

Renal and urinary system: renal impairment, renal failure

Respiratory, thoracic and mediastinal disorders: dyspnea

Skin and subcutaneous tissue disorders: urticaria

Vascular disorders: venous thrombosis

Nosocomial Pneumonia, including Ventilator-associated Pneumonia

ZERBAXA was evaluated in a Phase 3 comparator-controlled clinical trial for nosocomial pneumonia, which included a total of 361 patients treated with ZERBAXA (3 g every 8 hours, adjusted based on renal function where appropriate) and 359 patients treated with comparator (meropenem 1 g every

8 hours) for up to 14 days. The mean age of treated patients was 60 years (range 18 to 98 years), across treatment arms. About 44% of the subjects were 65 years of age or older. Most patients (71%) enrolled in the trial were male. All subjects were mechanically ventilated and 92% were in an intensive care unit (ICU) at randomization. The median APACHE II score was 17 and 33% of subjects had a baseline APACHE II score of ≥ 20 , indicating a high severity of illness for many patients enrolled in this trial. Table 6 lists adverse reactions occurring in 2% or greater of patients receiving ZERBAXA in a Phase 3 nosocomial pneumonia clinical trial.

Table 6: Adverse Reactions Occurring in 2% or Greater of Patients Receiving ZERBAXA in a Phase 3 Nosocomial Pneumonia Clinical Trial

Preferred Term	Nosocomial Pneumonia, including Ventilator-associated Pneumonia	
	ZERBAXA* N=361 n (%)	Meropenem N=359 n (%)
Gastrointestinal disorders		
Diarrhea	23 (6.4)	25 (7.0)
Vomiting	12 (3.3)	10 (2.8)
Infections and Infestations		
<i>Clostridium difficile</i> colitis ¹	10 (2.8)	2 (0.6)
Investigations		
ALT increased	21 (5.8)	14 (3.9)
AST increased	19 (5.3)	14 (3.9)
Transaminases increased	11 (3.0)	10 (2.8)
Vascular disorders		
Intracranial hemorrhage ²	16 (4.4)	5 (1.4)

*The ZERBAXA for injection dose was 3 g intravenously every 8 hours, adjusted to match renal function where appropriate.

¹Includes *Clostridium difficile* colitis, *Clostridium difficile* infection, *Clostridium* test positive.

²Includes cerebellar hemorrhage, cerebral hematoma, cerebral hemorrhage, hemorrhage intracranial, hemorrhagic stroke, hemorrhagic transformation stroke, intraventricular hemorrhage, subarachnoid hemorrhage, subdural hematoma.

Treatment discontinuation due to treatment-related adverse events occurred in 1.1% (4/361) of patients receiving ZERBAXA and 1.4% (5/359) of patients receiving meropenem.

Less Common Adverse Reactions in a Phase 3 Nosocomial Pneumonia Clinical Trial

The following selected adverse reactions were reported in ZERBAXA-treated subjects at a rate of less than 2%:

Infections and infestations: Clostridium difficile infection

Investigations: liver function test abnormal, blood alkaline phosphatase increased, gamma-glutamyltransferase increased, *Clostridium* test positive, Coombs direct test positive

Laboratory Values

The development of a positive direct Coombs test may occur during treatment with ZERBAXA. The incidence of seroconversion to a positive direct Coombs test was 0.2% in patients receiving ZERBAXA and 0% in patients receiving the comparator in the cUTI and cIAI clinical trials. The incidence of seroconversion to a positive direct Coombs test was 31.2% in patients receiving ZERBAXA and 3.6% in patients receiving meropenem in the nosocomial pneumonia clinical trial. In clinical studies, there was no evidence of hemolysis in patients who developed a positive direct Coombs test in any treatment group.

7 USE IN SPECIFIC POPULATIONS

7.1 Pregnancy

Pregnancy Category B.

There are no adequate and well-controlled trials in pregnant women with either ceftolozane or tazobactam. Because animal reproduction studies are not always predictive of human response, ZERBAXA should be used during pregnancy only if the potential benefit outweighs the possible risk.

Embryo-fetal development studies performed with intravenous ceftolozane in mice and rats with doses up to 2000 and 1000 mg/kg/day, respectively, revealed no evidence of harm to the fetus. The mean plasma exposure (AUC) values associated with these doses are approximately 3.5 (mice) and 2 (rats) times the mean daily human ceftolozane exposure at the highest recommended human dose of 2 grams every 8 hours. It is not known if ceftolozane crosses the

placenta in animals.

In a pre-postnatal study in rats, intravenous ceftolozane administered during pregnancy and lactation (Gestation Day 6 through Lactation Day 20) was associated with a decrease in auditory startle response in postnatal Day 60 male pups at maternal doses of greater than or equal to 300 mg/kg/day. The plasma exposure (AUC) associated with the NOAEL dose of 100 mg/kg/day in rats is lower than the mean daily human ceftolozane exposure at the highest recommended human dose of 2 grams every 8 hours.

In an embryo-fetal study in rats, tazobactam administered intravenously at doses up to 3000 mg/kg/day (approximately 10 times the highest recommended human dose of 1 gram every 8 hours based on body surface area comparison) produced maternal toxicity (decreased food consumption and body weight gain) but was not associated with fetal toxicity. In rats, tazobactam was shown to cross the placenta. Concentrations in the fetus were less than or equal to 10% of those found in maternal plasma.

In a pre-postnatal study in rats, tazobactam administered intraperitoneally twice daily at the end of gestation and during lactation (Gestation Day 17 through Lactation Day 21) produced decreased maternal food consumption and body weight gain at the end of gestation and significantly more stillbirths with a tazobactam dose of 1280 mg/kg/day (approximately 4 times the highest recommended human dose of 1 gram every 8 hours based on body surface area comparison). No effects on the development, function, learning or fertility of F1 pups were noted, but postnatal body weights for F1 pups delivered to dams receiving 320 and 1280 mg/kg/day tazobactam were significantly reduced 21 days after delivery. F2-generation fetuses were normal for all doses of tazobactam. The NOAEL for reduced F1 body weights was considered to be 40 mg/kg/day, a dose lower than the highest recommended human dose of 1 gram every 8 hours based on body surface area comparison.

7.2 Nursing Mothers

It is not known whether ceftolozane or tazobactam is excreted in human milk. Because many drugs are excreted in human milk, exercise caution when administering ZERBAXA to a nursing woman.

7.3 Pediatric Use

Safety and effectiveness in pediatric patients have not been established.

7.4 Geriatric Use

Of the 1015 patients treated with ZERBAXA in the Phase 3 cIAI and cUTI clinical trials, 250 (24.6%) were 65 years or older, including 113 (11.1%) 75 years or older. The incidence of adverse events in both treatment groups was higher in older subjects (65 years or older) in the trials for both indications. In the cIAI trial, cure rates in the elderly (aged 65 years and older) in the ZERBAXA plus metronidazole arm were 69/100 (69%) and in the comparator arm were 70/85 (82.4%). This finding in the elderly population was not observed in the cUTI trial.

Of the 361 patients treated with ZERBAXA in the Phase 3 nosocomial pneumonia clinical trial, 160 (44.3%) were 65 years or older, including 83 (23%) 75 years or older. The incidence of adverse events in both treatment groups was higher in older subjects (65 years or older). In the trial, Day 28 all-cause mortality rates in the elderly (aged 65 years and older) were comparable between treatment arms: 50/160 (31.3%) in the ZERBAXA arm and 54/160 (33.8%) in the comparator arm.

ZERBAXA is substantially excreted by the kidney and the risk of adverse reactions to ZERBAXA may be greater in patients with impaired renal function. Because elderly patients are more likely to have decreased renal function, care should be taken in dose selection and it may be useful to monitor renal function. Adjust dosage for elderly patients based on renal function [*see Dosage and Administration (2.2)*].

7.5 Patients with Renal Impairment

Dosage adjustment is required in patients with moderate (CrCl 30 to 50 mL/min) or severe (CrCl 15 to 29 mL/min) renal impairment and in patients with ESRD on HD [*see Dosage and Administration (2.2)*, *Warnings and Precautions (5.1)* and *Clinical Pharmacology (10.3)*].

8 OVERDOSAGE

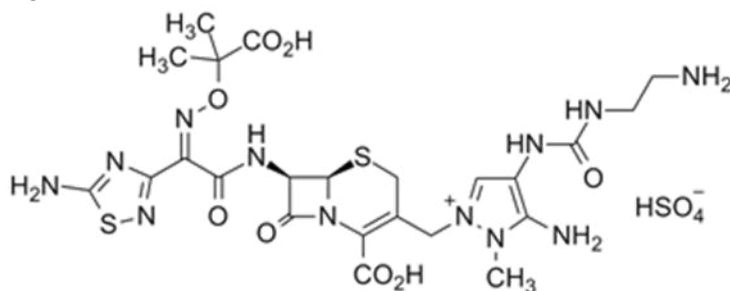
In the event of overdose, discontinue ZERBAXA and provide general supportive treatment. ZERBAXA can be removed by hemodialysis. Approximately 66% of ceftolozane, 56% of tazobactam, and 51% of the tazobactam metabolite M1 were removed by dialysis. No information is available on the use of hemodialysis to treat overdose.

9 DESCRIPTION

ZERBAXA (ceftolozane and tazobactam) is an antibacterial combination product consisting of the cephalosporin antibacterial drug ceftolozane sulfate and the beta-lactamase inhibitor tazobactam sodium for intravenous administration.

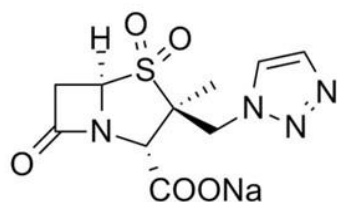
Ceftolozane sulfate is a semi-synthetic antibacterial drug of the beta-lactam class for parenteral administration. The chemical name of ceftolozane sulfate is 1*H*-Pyrazolium, 5-amino-4-[[[(2-aminoethyl)amino]carbonyl]amino]-2-[[[(6*R*,7*R*)-7-[[[(2*Z*)-2-(5-amino-1,2,4-thiadiazol-3-yl)-2-[(1-carboxy-1-methylethoxy)imino]acetyl]amino]-2-carboxy-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-en-3-yl]methyl]-1-methyl-,sulfate (1:1). The molecular formula is $C_{23}H_{31}N_{12}O_8S_2^+ \cdot HSO_4^-$ and the molecular weight is 764.77.

Figure 1: Chemical structure of ceftolozane sulfate



Tazobactam sodium, a derivative of the penicillin nucleus, is a penicillanic acid sulfone. Its chemical name is sodium (2*S*,3*S*,5*R*)-3-methyl-7-oxo-3-(1*H*-1,2,3-triazol-1-ylmethyl)-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylate-4,4-dioxide. The chemical formula is $C_{10}H_{11}N_4NaO_5S$ and the molecular weight is 322.3.

Figure 2: Chemical structure of tazobactam sodium



ZERBAXA 1.5 g (ceftolozane and tazobactam) for injection is a white to yellow sterile powder for reconstitution consisting of ceftolozane 1 g (equivalent to 1.147 g of ceftolozane sulfate) and tazobactam 0.5 g (equivalent to 0.537 g of tazobactam sodium) per vial, packaged in single-dose glass vials. The product contains sodium chloride (487 mg/vial) as a stabilizing agent, citric acid (21 mg/vial), and L-arginine (approximately 600 mg/vial) as excipients.

10 CLINICAL PHARMACOLOGY

10.1 Mechanism of Action

ZERBAXA is an antibacterial drug [see *Clinical Pharmacology* (10.4)].

10.2 Pharmacodynamics

As with other beta-lactam antibacterial agents, the time that the plasma concentration of ceftolozane exceeds the minimum inhibitory concentration (MIC) of the infecting organism has been shown to be the best predictor of efficacy in animal models of infection. The time above a threshold concentration has been determined to be the parameter that best predicts the efficacy of tazobactam in *in vitro* and *in vivo* nonclinical models. The exposure-response analyses in efficacy and safety clinical trials for cIAI, cUTI, and nosocomial pneumonia support the recommended dose regimens of ZERBAXA.

Cardiac Electrophysiology

In a randomized, positive and placebo-controlled crossover thorough QTc study, 51 healthy subjects were administered a single therapeutic dose of ZERBAXA 1.5 gram (ceftolozane 1 g and tazobactam 0.5 g) and a supratherapeutic dose of ZERBAXA 4.5 gram (ceftolozane 3 g and tazobactam 1.5 g). No significant effects of ZERBAXA on heart rate, electrocardiogram morphology, PR, QRS, or QT interval were detected. Therefore, ZERBAXA does not affect cardiac repolarization.

10.3 Pharmacokinetics

The mean pharmacokinetic parameters of ZERBAXA in healthy adults with normal renal function after multiple 1-hour intravenous infusions of ZERBAXA 1.5 g (ceftolozane 1 g and tazobactam 0.5 g) or 3 g (ceftolozane 2 g and tazobactam 1 g) administered every 8 hours are summarized in Table 7. Ceftolozane and tazobactam pharmacokinetics are similar following single- and multiple-dose administrations. The C_{max} and AUC of ceftolozane and tazobactam increase in proportion to dose. The elimination half-life ($t_{1/2}$) of ceftolozane or tazobactam is independent of dose.

Table 7: Mean (CV%) Steady-State Plasma Pharmacokinetic Parameters of ZERBAXA (ceftolozane and tazobactam) After Multiple Intravenous 1-hour Infusions of ZERBAXA 1.5 g (ceftolozane 1 g and tazobactam 0.5 g) or 3 g (ceftolozane 2 g and tazobactam 1 g) Every 8 Hours in Healthy Adults

PK parameters	ZERBAXA 1.5 g (ceftolozane 1 g and tazobactam 0.5 g)		ZERBAXA 3 g (ceftolozane 2 g and tazobactam 1 g)	
	Ceftolozane (n=10)	Tazobactam (n=10)	Ceftolozane (n=7)	Tazobactam (n=7)
C _{max} (mcg/mL)	74.4 (14)	18.0 (8)	112 (13)	25.8 (15)
t _{max} (h) [†]	1.07 (1.00, 1.10)	1.01 (1.00, 1.10)	1.0 (1.0, 1.0)	1.0 (0.5, 1.0)
AUC _{0-8,ss} (mcg•h/mL) [‡]	182 (15)	25.0 (15)	300 (9.8)	40.5 (13)
t _{1/2} (h)	3.12 (22)	1.03 (19)	2.8 (14)	1.0 (18)

[†] Median (minimum, maximum)

[‡] Steady-state AUC for 8 hour dosing interval. Daily AUC at steady state is calculated by multiplying the AUC_{0-8,ss} values by three (e.g., 546 mcg•h/mL for ceftolozane and 75 mcg•h/mL for tazobactam at the ceftolozane 1 g and tazobactam 0.5 g dosing regimen)

The mean steady-state population pharmacokinetic parameters of ZERBAXA in patients with cIAI and cUTI receiving 1 hour intravenous infusion of ZERBAXA 1.5 g (ceftolozane 1 g and tazobactam 0.5 g) or patients with nosocomial pneumonia receiving 1 hour intravenous infusion of ZERBAXA 3 g (ceftolozane 2 g and tazobactam 1 g) every 8 hours are summarized in Table 8.

Table 8: Mean (CV%) Steady-State Plasma Population Pharmacokinetic Parameters of ZERBAXA (ceftolozane and tazobactam) After Multiple Intravenous 1 hour Infusions of ZERBAXA 1.5 g (ceftolozane 1 g and tazobactam 0.5 g) or 3 g (ceftolozane 2 g and tazobactam 1 g) Every 8 Hours in Patients with CrCL greater than 50 mL/min

PK parameters	ZERBAXA 1.5 g (ceftolozane 1 g and tazobactam 0.5 g) in cIAI and cUTI Patients		ZERBAXA 3 g (ceftolozane 2 g and tazobactam 1 g) in Nosocomial Pneumonia Patients	
	Ceftolozane (n=317)	Tazobactam (n=244)	Ceftolozane (n=247)	Tazobactam (n=247)

$C_{\max}$ (mcg/mL)	65.7 (41)	17.8 (51)	105 (44)	26.4 (49)
$AUC_{0-8,ss}$ (mcg•h/mL)	186 (40)	35.8 (160)	392 (60)	73.3 (104)
$t_{1/2}$ (h)	2.7 (32)	1.8 (83)	3.9 (50)	3.2 (61)

Distribution

The binding of ceftolozane and tazobactam to human plasma proteins is approximately 16% to 21% and 30%, respectively. The mean (CV%) steady-state volume of distribution of ZERBAXA in healthy adult males (n = 51) following a single intravenous dose of ZERBAXA 1.5 g (ceftolozane 1 g and tazobactam 0.5 g) was 13.5 L (21%) and 18.2 L (25%) for ceftolozane and tazobactam, respectively, similar to extracellular fluid volume.

Following 1 hour intravenous infusions of ZERBAXA 3 g (ceftolozane 2 g and tazobactam 1 g) or adjusted based on renal function every 8 hours in ventilated patients with confirmed or suspected pneumonia (N=22), ceftolozane and tazobactam concentrations in pulmonary epithelial lining fluid were greater than 8 mcg/mL and 1 mcg/mL, respectively, over 100% of the dosing interval. Mean pulmonary epithelial-to-free plasma AUC ratios of ceftolozane and tazobactam were approximately 50% and 62%, respectively and are similar to those in healthy subjects (approximately 61% and 63%, respectively) receiving ZERBAXA 1.5 g (ceftolozane 1 g and tazobactam 0.5 g).

Elimination

Ceftolozane is eliminated from the body by renal excretion with a half-life of approximately 3 hours. Tazobactam is eliminated by renal excretion and metabolism with a plasma half-life of approximately 1 hour.

Metabolism

Ceftolozane is not a substrate for CYP enzymes and is mainly eliminated in the urine as unchanged parent drug and thus does not appear to be metabolized to any appreciable extent. The beta-lactam ring of tazobactam is hydrolyzed to form the pharmacologically inactive tazobactam metabolite M1.

Excretion

Ceftolozane, tazobactam and the tazobactam metabolite M1 are excreted by the kidneys.

Following administration of a single ZERBAXA 1.5 g (ceftolozane 1 g and tazobactam 0.5 g) intravenous dose to healthy male adults, greater than 95% of ceftolozane was excreted in the urine as unchanged parent drug. More than 80% of tazobactam was excreted as the parent compound with the remainder excreted as the tazobactam M1 metabolite. After a single dose of ZERBAXA, renal clearance of ceftolozane (3.41 – 6.69 L/h) was similar to plasma CL (4.10 to 6.73 L/h) and similar to the glomerular filtration rate for the unbound fraction, suggesting that ceftolozane is eliminated by the kidney via glomerular filtration. Tazobactam is a substrate for OAT1 and OAT3 transporters and its elimination has been shown to be inhibited by probenecid, an inhibitor of OAT1/3.

Specific Populations

Patients with Renal Impairment

ZERBAXA and the tazobactam metabolite M1 are eliminated by the kidneys.

The ceftolozane dose normalized geometric mean AUC increased up to 1.26-fold, 2.5-fold, and 5-fold in subjects with mild, moderate, and severe renal impairment, respectively, compared to healthy subjects with normal renal function. The respective tazobactam dose normalized geometric mean AUC increased approximately up to 1.3-fold, 2-fold, and 4-fold. To maintain similar systemic exposures to those with normal renal function, dosage adjustment is required [*see Dosage and Administration (2.2)*].

In subjects with ESRD on HD, approximately two-thirds of the administered ZERBAXA dose is removed by HD. The recommended dose in cIAI or cUTI subjects with ESRD on HD is a single loading dose of ZERBAXA 750 mg (ceftolozane 500 mg and tazobactam 250 mg), followed by a ZERBAXA 150 mg (ceftolozane 100 mg and tazobactam 50 mg) maintenance dose administered every 8 hours for the remainder of the treatment period. The recommended dose in nosocomial pneumonia subjects with ESRD on HD is a single loading dose of ZERBAXA 2.25 g (ceftolozane 1.5 g and tazobactam 0.75 g), followed by a ZERBAXA 450 mg (ceftolozane 300 mg and tazobactam 150 mg) maintenance dose administered every 8 hours for the remainder of the treatment period. On HD days, administer the dose at the earliest possible time following completion of HD [*see Dosage and Administration (2.2)*].

Augmented renal clearance

Following a single 1 hour intravenous infusion of ZERBAXA 3 g (ceftolozane 2 g and tazobactam

1 g) to critically ill patients with CrCL greater than or equal to 180 mL/min (N=10), mean terminal half-life values of ceftolozane and tazobactam were 2.6 hours and 1.5 hours, respectively. Free plasma ceftolozane concentrations were greater than 8 mcg/mL over 70% of an 8-hour period; free tazobactam concentrations were greater than 1 mcg/mL over 60% of an 8-hour period. No dose adjustment of ZERBAXA is recommended for nosocomial pneumonia patients with augmented renal clearance [see Clinical Studies (12.3)].

Patients with Hepatic Impairment

As ZERBAXA does not undergo hepatic metabolism, the systemic clearance of ZERBAXA is not expected to be affected by hepatic impairment.

No dose adjustment is recommended for ZERBAXA in subjects with hepatic impairment.

Geriatric Patients

In a population pharmacokinetic analysis of ZERBAXA, no clinically relevant differences in exposure were observed with regard to age.

No dose adjustment of ZERBAXA based on age is recommended. Dosage adjustment for ZERBAXA in elderly patients should be based on renal function [see Dosage and Administration (2.2)].

Pediatric Patients

Safety and effectiveness in pediatric patients have not been established.

Gender

In a population pharmacokinetic analysis of ZERBAXA, no clinically relevant differences in AUC were observed for ceftolozane and tazobactam.

No dose adjustment is recommended based on gender.

Race

In a population pharmacokinetic analysis of ZERBAXA, no clinically relevant differences in ZERBAXA AUC were observed in Caucasians compared to other races combined.

No dose adjustment is recommended based on race.

Drug Interactions

No drug-drug interaction was observed between ceftolozane and tazobactam in a clinical study in 16 healthy subjects. *In vitro* and *in vivo* data indicate that ZERBAXA is unlikely to cause clinically relevant drug-drug interactions related to CYPs and transporters at therapeutic concentrations.

Drug Metabolizing Enzymes

In vivo data indicated that ZERBAXA is not a substrate for CYPs. Thus clinically relevant drug-drug interactions involving inhibition or induction of CYPs by other drugs are unlikely to occur.

In vitro studies demonstrated that ceftolozane, tazobactam and the M1 metabolite of tazobactam did not inhibit CYP1A2, CYP2B6, CYP2C8, CYP2C9, CYP2C19, CYP2D6, or CYP3A4 and did not induce CYP1A2, CYP2B6, or CYP3A4 at therapeutic plasma concentrations. *In vitro* induction studies in primary human hepatocytes demonstrated that ceftolozane, tazobactam, and the tazobactam metabolite M1 decreased CYP1A2 and CYP2B6 enzyme activity and mRNA levels in primary human hepatocytes as well as CYP3A4 mRNA levels at supratherapeutic plasma concentrations. Tazobactam metabolite M1 also decreased CYP3A4 activity at supratherapeutic plasma concentrations. A clinical drug-drug interaction study was conducted and results indicated drug interactions involving CYP1A2 and CYP3A4 inhibition by ZERBAXA are not anticipated.

Membrane Transporters

Ceftolozane and tazobactam were not substrates for P-gp or BCRP, and tazobactam was not a substrate for OCT2, *in vitro* at therapeutic concentrations.

Tazobactam is a known substrate for OAT1 and OAT3. Co-administration of tazobactam with the OAT1/OAT3 inhibitor probenecid has been shown to prolong the half-life of tazobactam by 71%. Co-administration of ZERBAXA with drugs that inhibit OAT1 and/or OAT3 may increase tazobactam plasma concentrations.

In vitro data indicate that ceftolozane did not inhibit P-gp, BCRP, OATP1B1, OATP1B3, OCT1, OCT2, MRP, BSEP, OAT1, OAT3, MATE1, or MATE2-K *in vitro* at therapeutic plasma concentrations.

In vitro data indicate that neither tazobactam nor the tazobactam metabolite M1 inhibit P-gp, BCRP, OATP1B1, OATP1B3, OCT1, OCT2, or BSEP transporters at therapeutic plasma concentrations. *In vitro*, tazobactam inhibited human OAT1 and OAT3 transporters with IC₅₀ values of 118 and 147 mcg/mL, respectively. A clinical drug-drug interaction study was conducted and results

indicated clinically relevant drug interactions involving OAT1/OAT3 inhibition by ZERBAXA are not anticipated.

10.4 Microbiology

Mechanism of Action

Ceftolozane belongs to the cephalosporin class of antibacterial drugs. The bactericidal action of ceftolozane results from inhibition of cell wall biosynthesis, and is mediated through binding to penicillin-binding proteins (PBPs). Ceftolozane is an inhibitor of PBPs of *P. aeruginosa* (e.g., PBP1b, PBP1c, and PBP3) and *E. coli* (e.g., PBP3).

Tazobactam sodium has little clinically relevant *in vitro* activity against bacteria due to its reduced affinity to penicillin-binding proteins. It is an irreversible inhibitor of some beta-lactamases (e.g., certain penicillinases and cephalosporinases), and can bind covalently to some chromosomal and plasmid-mediated bacterial beta-lactamases.

In the 2017 Program to Assess Ceftolozane/Tazobactam Susceptibility (PACTS) surveillance study the overall ceftolozane/tazobactam susceptibility of 3937 Enterobacteriaceae isolates collected from all sources from US hospitals was 95.6% and against extended spectrum beta-lactamase (ESBL), non-carbapenem resistant Enterobacteriaceae isolates the percent ceftolozane/tazobactam susceptibility was 93.5%. The overall ceftolozane/tazobactam susceptibility of 910 *P. aeruginosa* isolates collected from US hospitals was 97.7%. When ceftolozane/tazobactam was tested against isolates non-susceptible to ceftazidime, meropenem or piperacillin/tazobactam, the percent susceptibility to ceftolozane/tazobactam was 87.2%, 91.3% and 89.5%, respectively.

Resistance

Mechanisms of beta-lactam resistance may include the production of beta-lactamases, modification of PBPs by gene acquisition or target alteration, up-regulation of efflux pumps, and loss of outer membrane porin.

Clinical isolates may produce multiple beta-lactamases, express varying levels of beta-lactamases, or have amino acid sequence variations, and other resistance mechanisms that have not been identified.

Culture and susceptibility information and local epidemiology should be considered in selecting

or modifying antibacterial therapy.

ZERBAXA demonstrated *in vitro* activity against Enterobacteriaceae in the presence of some extended-spectrum beta-lactamases (ESBLs) and other beta-lactamases of the following groups: TEM, SHV, CTX-M, and OXA. ZERBAXA is not active against bacteria that produce serine carbapenemases [*K. pneumoniae* carbapenemase (KPC)], and metallo-beta-lactamases.

In ZERBAXA clinical trials, some isolates of Enterobacteriaceae that produced beta-lactamases, were susceptible to ZERBAXA (minimum inhibitory concentration ≤ 2 mcg/mL). These isolates produced one or more beta-lactamases of the following enzyme groups: CTX-M, OXA, TEM, or SHV.

Some of these beta-lactamases were also produced by isolates of Enterobacteriaceae that were not susceptible to ZERBAXA (minimum inhibitory concentration > 2 mcg/mL). These isolates produced one or more beta-lactamases of the following enzyme groups: CTX-M, OXA, TEM, or SHV.

ZERBAXA demonstrated *in vitro* activity against *P. aeruginosa* isolates tested that had chromosomal AmpC, loss of outer membrane porin (OprD), or up-regulation of efflux pumps (MexXY, MexAB).

Isolates resistant to other cephalosporins may be susceptible to ZERBAXA, although cross-resistance may occur.

Interaction with Other Antimicrobials

In vitro synergy studies suggest no antagonism between ZERBAXA and other antibacterial drugs (e.g., meropenem, amikacin, aztreonam, levofloxacin, tigecycline, rifampin, linezolid, daptomycin, vancomycin, and metronidazole).

Antimicrobial Activity

ZERBAXA has been shown to be active against the following bacteria, both *in vitro* and in clinical infections [see *Indications and Usage (1)*].

Complicated Intra-abdominal Infections

Gram-negative bacteria:

Enterobacter cloacae

Escherichia coli

Klebsiella oxytoca

Klebsiella pneumoniae

Proteus mirabilis

Pseudomonas aeruginosa

Gram-positive bacteria:

Streptococcus anginosus

Streptococcus constellatus

Streptococcus salivarius

Anaerobic bacteria:

Bacteroides fragilis

Complicated Urinary Tract Infections, Including Pyelonephritis

Gram-negative bacteria:

Escherichia coli

Klebsiella pneumoniae

Proteus mirabilis

Pseudomonas aeruginosa

Nosocomial Pneumonia, including Ventilator-associated Pneumonia

Gram-negative bacteria

Enterobacter cloacae

Escherichia coli

Haemophilus influenzae

Klebsiella oxytoca

Klebsiella pneumoniae

Proteus mirabilis

Pseudomonas aeruginosa

Serratia marcescens

The following *in vitro* data are available, but their clinical significance is unknown. At least 90 percent of the following bacteria exhibit an *in vitro* minimum inhibitory concentration (MIC) less than or equal to the susceptible breakpoint for ceftolozane and tazobactam against isolates of similar genus or organism group. However, the efficacy of ZERBAXA in treating clinical infections due to these bacteria has not been established in adequate and well-controlled clinical trials.

Gram-negative bacteria:

Citrobacter koseri

Morganella morganii

Proteus vulgaris

Providencia rettgeri

Providencia stuartii

Serratia liquefaciens

Klebsiella (Enterobacter) aerogenes

Gram-positive bacteria:

Streptococcus agalactiae

Streptococcus intermedius

Susceptibility Test Methods

When available, the clinical microbiology laboratory should provide cumulative reports of *in vitro* susceptibility test results for antimicrobial drugs used in local hospitals and practice areas to the physician as periodic reports that describe the susceptibility profile of nosocomial and community-acquired pathogens. These reports should aid the physician in selecting an antibacterial drug for treatment.

Dilution Techniques:

Quantitative methods are used to determine antimicrobial minimum inhibitory concentrations (MICs). Ceftolozane and tazobactam susceptibility testing is performed with a fixed 4 mcg/mL concentration of tazobactam. These MICs provide estimates of the susceptibility of bacteria to antibacterial compounds. The MICs should be determined using a standardized test method (broth, and/or agar).^{1,4} The MIC values should be interpreted according to the criteria in Table 9.

Diffusion Techniques:

Quantitative methods that require measurement of zone diameters can also provide reproducible estimates of the susceptibility of bacteria to antimicrobial compounds. The zone size should be determined using a standardized test method.^{2,4} This procedure uses paper disks impregnated with 30 mcg of ceftolozane and 10 mcg of tazobactam to test the susceptibility of bacteria to ceftolozane and tazobactam. The disk diffusion should be interpreted according to the criteria in Table 9.

Anaerobic Techniques:

For anaerobic bacteria, the susceptibility to ceftolozane and tazobactam can be determined by standardized test method.³ The MIC values obtained should be interpreted according to criteria provided in Table 9.

Table 9: Susceptibility Test Interpretive Criteria for Ceftolozane/Tazobactam

Pathogen	Minimum Inhibitory Concentrations (mcg/mL)			Disk Diffusion Zone Diameter (mm)		
	S	I	R	S	I	R
Enterobacteriaceae	≤2/4	4/4	≥8/4	≥22	19-21	≤18
<i>Pseudomonas aeruginosa</i>	≤4/4	8/4	≥16/4	≥21	17-20	≤16
<i>Haemophilus influenzae</i> (nosocomial pneumonia, including VAP) [†]	≤0.5/4	---	---	---	---	---
<i>Streptococcus anginosus</i> <i>Streptococcus constellatus</i> and <i>Streptococcus salivarius</i> (cIAI and cUTI, including pyelonephritis)*	≤8/4	16/4	≥32/4	---	---	---
<i>Bacteroides fragilis</i> (cIAI and cUTI, including pyelonephritis)*	≤8/4	16/4	≥32/4	---	---	---

S = susceptible, I = intermediate, R = resistant

*Based on 1.5 g IV every 8 hours

†Based on 3 g IV every 8 hours

A report of *Susceptible (S)* indicates that the antimicrobial is likely to inhibit growth of the pathogen if the antimicrobial drug reaches the concentration usually achievable at the site of infection. A report of *Intermediate (I)* indicates that the result should be considered equivocal, and if the microorganism is not fully susceptible to alternative clinically feasible drugs, the test should be repeated. This category implies possible clinical applicability in body sites where the drug is physiologically concentrated or in situations where a high dose of the drug can be used. This category also provides a buffer zone that prevents small uncontrolled technical factors from causing major discrepancies in interpretation. A report of *Resistant (R)* indicates that the antimicrobial is not likely to inhibit growth of the pathogen if the antimicrobial drug reaches the concentrations usually achievable at the infection site; other therapy should be selected.

Quality Control

Standardized susceptibility test procedures require the use of laboratory controls to monitor and ensure the accuracy and precision of supplies and reagents used in the assay, and the techniques of the individuals performing the test.^{1,2,3,4} Standard ceftolozane and tazobactam powder should provide the following range of MIC values provided in Table 10. For the diffusion technique using the 30 mcg ceftolozane/10 mcg tazobactam disk, the criteria provided in Table 10 should be achieved.⁴

Table 10: Acceptable Quality Control Ranges for Ceftolozane/Tazobactam

Quality Control Strain	Minimum Inhibitory Concentrations (mcg/mL)	Disk Diffusion (Zone Diameters in mm)
<i>Escherichia coli</i> ATCC 25922	0.12/4-0.5/4	24-32
<i>Escherichia coli</i> * ATCC 35218	0.06/4-0.25/4	25-31
<i>Pseudomonas aeruginosa</i> ATCC 27853	0.25/4-1/4	25-31
<i>Staphylococcus aureus</i> ATCC 25923	Not Applicable	10-18
<i>Staphylococcus aureus</i> ATCC 29213	16/4-64/4	Not Applicable

<i>Haemophilus influenzae</i> [†] ATCC 49247	0.5/4-2/4	23-29
<i>Klebsiella pneumoniae</i> * ATCC 700603	0.5/4-2/4	17-25
<i>Streptococcus pneumoniae</i> ATCC 49619	0.25/4-1/4	21-29
<i>Bacteroides fragilis</i> ATCC 25285 (agar and broth)	0.12/4-1/4	Not Applicable
<i>Bacteroides thetaiotaomicron</i> ATCC 29741 (agar)	16/4-128/4	Not Applicable
<i>Bacteroides thetaiotaomicron</i> ATCC 29741 (broth)	16/4-64/4	Not Applicable

ATCC = American Type Culture Collection

* Store *E. coli* ATCC 35218 and *K. pneumoniae* ATCC 700603 stock cultures at -60°C or below and prepare working stock cultures weekly.

[†] This strain may lose its plasmid and develop susceptibility to beta-lactam antimicrobial agents after repeated transfers onto culture media. Minimize by removing new culture from storage at least monthly or whenever the strain begins to show increased zone diameters to ampicillin, piperacillin, or ticarcillin.

11 NONCLINICAL TOXICOLOGY

11.1 Carcinogenesis, Mutagenesis, Impairment of Fertility

Long-term carcinogenicity studies in animals have not been conducted with ZERBAXA, ceftolozane, or tazobactam.

ZERBAXA was negative for genotoxicity in an *in vitro* mouse lymphoma assay and an *in vivo* rat bone-marrow micronucleus assay. In an *in vitro* chromosomal aberration assay in Chinese hamster ovary cells, ZERBAXA was positive for structural aberrations.

Ceftolozane was negative for genotoxicity in the *in vitro* microbial mutagenicity (Ames) assay, the *in vitro* chromosomal aberration assay in Chinese hamster lung fibroblast cells, the *in vitro* mouse lymphoma assay, the *in vitro* HPRT assay in Chinese hamster ovary cells, the *in vivo* mouse micronucleus assay, and the *in vivo* unscheduled DNA synthesis (UDS) assay.

Tazobactam was negative for genotoxicity in an *in vitro* microbial mutagenicity (Ames) assay, an *in vitro* chromosomal aberration assay in Chinese hamster lung cells, a mammalian point-mutation (Chinese hamster ovary cell HPRT) assay, an *in vivo* mouse bone-marrow micronucleus assay, and a UDS assay.

Ceftolozane had no adverse effect on fertility in male or female rats at intravenous doses up to 1000 mg/kg/day. The mean plasma exposure (AUC) value at this dose is approximately 1.4 times the mean daily human ceftolozane exposure value at the highest recommended human dose of 2 grams every 8 hours.

In a rat fertility study with intraperitoneal tazobactam twice-daily, male and female fertility parameters were not affected at doses less than or equal to 640 mg/kg/day (approximately 2 times the highest recommended human dose of 1 gram every 8 hours based on body surface comparison).

12 CLINICAL STUDIES

12.1 Complicated Intra-abdominal Infections

A total of 979 adults hospitalized with cIAI were randomized and received study medications in a multinational, double-blind study comparing ZERBAXA 1.5 g (ceftolozane 1 g and tazobactam 0.5 g) intravenously every 8 hours plus metronidazole (500 mg intravenously every 8 hours) to meropenem (1 g intravenously every 8 hours) for 4 to 14 days of therapy. Complicated intra-abdominal infections included appendicitis, cholecystitis, diverticulitis, gastric/duodenal perforation, perforation of the intestine, and other causes of intra-abdominal abscesses and peritonitis.

The primary efficacy endpoint was clinical response, defined as complete resolution or significant improvement in signs and symptoms of the index infection at the test-of-cure (TOC) visit which occurred 24 to 32 days after the first dose of study drug. The primary efficacy analysis population was the Clinically Evaluable (CE) population, which included all protocol adherent patients that received an adequate amount of study drug. The key secondary efficacy endpoint was clinical response at the TOC visit in the Intent-to-Treat (ITT) population, which included all randomized subjects regardless of whether or not the subjects went on to receive study drug.

The CE population consisted of 774 patients; the median age was 49 years and 58.7% were male. The most common diagnosis was appendiceal perforation or peri-appendiceal abscess, occurring in 47.7% of patients. Diffuse peritonitis at baseline was present in 35.9% of patients.

ZERBAXA plus metronidazole was non-inferior to meropenem with regard to clinical cure rates at the TOC visit in the CE population. Clinical cure rates at the TOC visit are displayed by patient population in Table 11. Clinical cure rates at the TOC visit by pathogen in the Microbiologically Evaluable (ME) population are presented in Table 12. The ME included all protocol adherent patients with at least 1 baseline intra-abdominal pathogen regardless of the susceptibility to study drug.

Table 11: Clinical Cure Rates in a Phase 3 Study of Complicated Intra-Abdominal Infections

Analysis Population	ZERBAXA plus metronidazole* n/N (%)	Meropenem† n/N (%)	Treatment Difference (99% CI)‡
CE Population	353/375 (94.1)	375/399 (94)	0 (-4.16, 4.3)
ITT Population	399/476 (83.8)	424/494 (85.8)	-2.2 (-7.95, 3.44)

* ZERBAXA 1.5 g IV every 8 hours + metronidazole 500 mg IV every 8 hours

† 1 g IV every 8 hours

‡ The 99% CI was calculated using the Newcombe method with minimum risk weights

Table 12: Per Pathogen Clinical Cure Rates in a Phase 3 Study of Complicated Intra-abdominal Infections (ME Population)

Organism Group Pathogen	ZERBAXA plus metronidazole n/N (%)	Meropenem n/N (%)
Aerobic Gram-negative	238/252 (94.4)	273/291 (93.8)
<i>Escherichia coli</i>	197/208 (94.7)	216/231 (93.5)
<i>Escherichia coli</i> (ESBL-producing)	14/14 (100)	18/20 (90)
<i>Escherichia coli</i> (CTX-M-14/15 ESBL-producing)	9/9 (100)	7/9 (77.8)
<i>Klebsiella pneumoniae</i>	28/30 (93.3)	22/25 (88)
<i>Klebsiella pneumoniae</i> (ESBL-producing)	7/8 (87.5)	3/4 (75)
<i>Klebsiella pneumoniae</i> (CTX-M-15 ESBL-producing)	5/5 (100)	0/1 (0)
<i>Pseudomonas aeruginosa</i>	26/26 (100)	27/29 (93.1)
<i>Enterobacter cloacae</i>	19/22 (86.4)	22/22 (100)
<i>Klebsiella oxytoca</i>	12/12 (100)	21/22 (95.5)

Organism Group Pathogen	ZERBAXA plus metronidazole n/N (%)	Meropenem n/N (%)
<i>Proteus mirabilis</i>	10/11 (90.9)	9/10 (90)
Aerobic Gram-positive	153/168 (91.1)	170/185 (91.9)
<i>Streptococcus anginosus</i>	25/30 (83.3)	23/23 (100)
<i>Streptococcus constellatus</i>	17/18 (94.4)	20/23 (87)
<i>Streptococcus salivarius</i>	9/10 (90)	8/8 (100)
Anaerobic Gram-negative	104/109 (95.4)	132/137 (96.4)
<i>Bacteroides fragilis</i>	39/41 (95.1)	56/57 (98.2)

In a subset of the *E. coli* and *K. pneumoniae* isolates from both arms of the cIAI Phase 3 trial that met pre-specified criteria for beta-lactam susceptibility, genotypic testing identified certain ESBL groups (e.g., TEM, SHV, CTX-M, OXA) in 53/601 (9%). Cure rates in this subset were similar to the overall trial results. *In vitro* susceptibility testing showed that some of these isolates were susceptible to ZERBAXA, while some others were not susceptible. Isolates of a specific genotype were seen in patients who were deemed to be either successes or failures.

12.2 Complicated Urinary Tract Infections, including Pyelonephritis

A total of 1068 adults hospitalized with complicated urinary tract infections (including pyelonephritis) were randomized and received study medications in a multinational, double-blind study comparing ZERBAXA (1.5 g IV every 8 hours) to levofloxacin (750 mg IV once daily) for 7 days of therapy. The primary efficacy endpoint was defined as microbiological eradication (all uropathogens found at baseline at $\geq 10^5$ were reduced to $< 10^3$ CFU/mL) at the test-of-cure (TOC) visit 7 (± 2) days after the last dose of study drug. The primary efficacy analysis population was the microbiologically evaluable (ME) population, which included protocol-adherent microbiologically modified intent-to-treat (mMITT) patients with a urine culture at the TOC visit. The key secondary efficacy endpoint was microbiological eradication at the TOC visit in the mMITT population, which included all patients who received study medication and had at least 1 baseline uropathogen.

The ME population consisted of 693 patients with cUTI, including 567 (82%) with pyelonephritis. The median age was 50 years and 73% were female. Concomitant bacteremia was identified in 50 (7.2%) patients at baseline.

ZERBAXA was superior to levofloxacin with regard to the microbiological eradication rates at the TOC visit in both the ME and mMITT populations (Table 13).

Microbiological eradication rates at the TOC visit by pathogen in the ME population are presented in Table 14.

Table 13: Microbiological Eradication Rates in a Phase 3 Study of Complicated Urinary Tract Infections

Analysis Population	ZERBAXA* n/N (%)	Levofloxacin† n/N (%)	Treatment Difference (99% CI)‡
ME	288/340 (84.7)	266/353 (75.4)	9.4 (1.54, 17.12)
mMITT	313/398 (78.6)	281/402 (69.9)	8.7 (0.77, 16.57)

* 1.5 g IV every 8 hours

† 750 mg IV once daily

‡ The 99% CI was based on the stratified Newcombe method

Table 14: Per Pathogen Microbiological Eradication Rates in a Phase 3 Study of Complicated Urinary Tract Infections (ME Population)

Organism Group Pathogen	ZERBAXA n/N (%)	Levofloxacin n/N (%)
Aerobic Gram-negative	282/322 (87.6)	255/340 (75)
<i>Escherichia coli</i>	232/261 (88.9)	219/284 (77.1)
<i>Escherichia coli</i> (ESBL-producing)	26/36 (72.2)	17/36 (47.2)
<i>Escherichia coli</i> (CTX-M-14/15 ESBL-producing)	19/27 (70.4)	13/25 (52)
<i>Klebsiella pneumoniae</i>	21/25 (84)	14/23 (60.9)
<i>Klebsiella pneumoniae</i> (ESBL-producing)	7/10 (70)	2/7 (28.6)
<i>Klebsiella pneumoniae</i> (CTX-M-15 ESBL-producing)	5/8 (62.5)	1/4 (25)
<i>Proteus mirabilis</i>	10/10 (100)	8/11 (72.7)
<i>Pseudomonas aeruginosa</i>	6/7 (85.7)	6/12 (50)

In patients with levofloxacin-resistant pathogens at baseline, ZERBAXA was superior to levofloxacin with regards to microbiological eradication rate in the ME population, 58/89 (65.2%) in the ZERBAXA treatment arm and 42/99 (42.4%) in the levofloxacin treatment arm (95% CI: 22.7 [8.47, 35.73]).

In the ME population, the microbiological eradication rate in patients with concurrent bacteremia were 21/24 (87.5%) for ZERBAXA and 20/26 (76.9%) for levofloxacin.

In a subset of the *E. coli* and *K. pneumoniae* isolates from both arms of the cUTI Phase 3 trial that met pre-specified criteria for beta-lactam susceptibility, genotypic testing identified certain ESBL groups (e.g., TEM, SHV, CTX-M, OXA) in 104/687 (15%). Cure rates in this subset were similar to the overall trial results. *In vitro* susceptibility testing showed that some of these isolates were susceptible to ZERBAXA, while some others were not susceptible. Isolates of a specific genotype were seen in patients who were deemed to be either successes or failures.

12.3 Nosocomial Pneumonia, including Ventilator-associated Pneumonia

A total of 726 adult patients hospitalized with ventilated nosocomial pneumonia (including hospital-acquired pneumonia and ventilator-associated pneumonia) were enrolled in a multinational, double-blind study comparing ZERBAXA 3 g (ceftolozane 2 g and tazobactam 1 g) intravenously every 8 hours to meropenem (1 g intravenously every 8 hours) for 8 to 14 days of therapy. All patients had to be intubated and on mechanical ventilation at randomization.

The primary efficacy endpoint was all-cause mortality at Day 28. Clinical response, defined as complete resolution or significant improvement in signs and symptoms of the index infection at the test-of-cure (TOC) visit which occurred 7 to 14 days after the end of treatment was a pre-specified key secondary endpoint. The analysis population for both the primary and key secondary endpoints was the intent-to-treat (ITT) population, which included all randomized patients.

Following a diagnosis of HABP/VABP and prior to receipt of first dose of study drug, if required, patients could have received up to a maximum of 24 hours of active non-study antibacterial drug therapy in the 72 hours preceding the first dose of study drug. Patients who had failed prior antibacterial drug therapy for the current episode of HABP/VABP could be enrolled if the baseline lower respiratory tract (LRT) culture showed growth of a Gram-negative pathogen while the patient was on the antibacterial therapy and all other eligibility criteria were met. Empiric therapy at baseline with linezolid or other approved therapy for Gram-positive coverage was required in all patients pending baseline LRT culture results. Adjunctive Gram-negative therapy was optional and allowed for a maximum of 72 hours in centers with a prevalence of meropenem-resistant *P. aeruginosa* more than 15%.

Of the 726 patients in the ITT population the median age was 62 years and 44% of the population

was greater than or equal to 65 years of age, with 22% of the population greater than or equal to 75 years of age. The majority of patients were white (83%), male (71%) and were from Eastern Europe (64%). The median APACHE II score was 17 and 33% of subjects had a baseline APACHE II score of greater than or equal to 20. All subjects were on mechanical ventilation and 519 (71%) had VAP. At randomization, the majority of subjects had been hospitalized for greater than or equal to 5 days (77%), ventilated for greater than or equal to 5 days (49%) and in an ICU (92%). Approximately 36% of patients had renal impairment at baseline and 14% had moderate or severe impairment (CrCL less than 50 mL/min). Approximately 13% of subjects had failed prior antibiotic treatment for nosocomial pneumonia and bacteremia was present at baseline in 15% of patients. Key comorbidities included chronic obstructive pulmonary disease (COPD), diabetes mellitus, and congestive heart failure at rates of 12%, 22% and 16%, respectively. In both treatment groups, most subjects (63.1%) received between 8 and 14 days of study therapy as specified in the protocol.

In the ITT population, Day 28 all-cause mortality and clinical cure rates in patients with CrCL greater than or equal to 150 mL/min were similar between ZERBAXA and meropenem. In patients with bacteremia at baseline, Day 28 all-cause mortality rates were 23/64 (35.9%) for ZERBAXA-treated patients and 13/41 (31.7%) for meropenem-treated patients; clinical cure rates were 30/64 (46.9%) and 15/41 (36.6%), respectively.

In the ITT population, ZERBAXA was non-inferior to meropenem with regard to the primary endpoint of all-cause mortality at Day 28 and key secondary endpoint of clinical cure rates at the TOC visit (Table 15).

Table 15: 28-Day All-cause Mortality and Clinical Cure Rates at TOC from a Phase 3 Study of Nosocomial Pneumonia (ITT Population)

Endpoint	ZERBAXA	Meropenem	Treatment Difference (95% CI) [‡]
Day 28 All-cause Mortality	87/362 (24.0)	92/364 (25.3)	1.1 (-5.13, 7.39)
VAP	63/263 (24.0)	52/256 (20.3)	-3.6 (-10.74, 3.52)
Ventilated HAP	24/99 (24.2)	40/108 (37.0)	12.8 (0.18, 24.75)
Clinical Cure at TOC Visit	197/362 (54.4)	194/364 (53.3)	1.1 (-6.17, 8.29)
VAP	147/263 (55.9)	146/256 (57.0)	-1.1 (-9.59, 7.35)
Ventilated HAP	50/99 (50.5)	48/108 (44.4)	6.1 (-7.44, 19.27)

[‡]The CI for overall treatment difference was based on the stratified Newcombe method with minimum

risk weights. The CI for treatment difference of each primary diagnosis was based on the unstratified Newcombe method.

In the ITT population, Day 28 all-cause mortality rates in patients with renal hyperclearance at baseline (CrCL greater than or equal to 150 mg/mL) were 10/67 (14.9%) for ZERBAXA and 7/64 (10.9%) for meropenem; the clinical cure rates were 40/67 (59.7%) and 39/64 (60.9%), respectively. In those patients who failed prior antibiotic therapy for nosocomial pneumonia, Day 28 all-cause mortality rates were 12/53 (22.6%) for ZERBAXA and 18/40 (45%) for meropenem; the clinical cure rates were 26/53 (49.1%) and 15/40 (37.5%), respectively. In patients with bacteremia at baseline, Day 28 all-cause mortality rates were 23/64 (35.9%) for ZERBAXA and 13/41 (31.7%) for meropenem; clinical cure rates were 30/64 (46.9%) and 15/41 (36.6%), respectively.

In the ventilated HABP sub-group, a favorable response for ZERBAXA in 28-day all-cause mortality was observed, 24.2% (24/99) for ZERBAXA and 37.0% (40/108) for meropenem, respectively, for a weighted proportion difference of 12.8 (stratified 95% CI: 0.18, 24.75). In the VABP subgroup, 28-day all-cause mortality was 24.0% (63/263) for ZERBAXA and 20.3% (52/256) for meropenem, for a weighted proportion difference of -3.6 (stratified 95% CI: -10.74, 3.52).

Per pathogen clinical and microbiologic responses were assessed in the microbiologic intention to treat population (mITT), which consisted of all randomized subjects who had a baseline lower respiratory tract (LRT) pathogen that was susceptible to at least one of the study therapies, and in the microbiologically evaluable (ME) population, which included protocol-adherent mITT patients with a baseline LRT pathogen that grew at the appropriate colony-forming unit (CFU)/mL threshold. In the mITT and ME populations, *Klebsiella pneumoniae* (34.6% and 38.6%, respectively) and *Pseudomonas aeruginosa* (25% and 28.8%, respectively) were the most prevalent pathogens isolated from baseline LRT cultures. Among all Enterobacteriaceae, 157 (30.7%) in the mITT and 84 (36.1%) in the ME were ESBL-positive; among all *K. pneumoniae* isolates, 105 (20.5%) in the mITT and 57 (24.5%) in the ME were ESBL-positive. AmpC-overexpression among *P. aeruginosa* was detected in 15 (2.9%) and 9 (3.9%) of the *P. aeruginosa* isolates in the mITT and ME populations, respectively. Clinical cure rates at TOC by pathogen in the mITT and ME populations are presented in Table 16. In the mITT population clinical cure rates in patients with a Gram-negative pathogen at baseline were 157/259 (60.6%) for ZERBAXA and 137/240 (57.1%) for meropenem; results were consistent in the ME population with 85/113 (75.2%) and 78/117 (66.7%) clinical cure rates, respectively. Microbiologic response rates at TOC by pathogen in the mITT and ME populations are presented in Table 17. In the mITT population microbiologic response rates in patients with a Gram-negative pathogen at baseline were 189/259

(73%) for ZERBAXA and 163/240 (67.9%) for meropenem; results were consistent in the ME population with 79/113 (69.9%) and 73/117 (62.4%) microbiologic response rates, respectively.

Table 16: Clinical Cure Rates by Baseline Pathogen from a Phase 3 Study of Nosocomial Pneumonia (mITT and ME populations)

Baseline Pathogen Category Baseline Pathogen	mITT Population		ME Population	
	ZERBAXA n/N (%)	Meropenem n/N (%)	ZERBAXA n/N (%)	Meropenem n/N (%)
<i>Pseudomonas aeruginosa</i>	36/63 (57.1)	39/65 (60.0)	23/29 (79.3)	28/38 (73.7)
AmpC Overexpressing <i>Pseudomonas aeruginosa</i>	4/9 (44.4)	3/6 (50.0)	2/4 (50.0)	3/5 (60.0)
Enterobacteriaceae	120/195 (61.5)	105/185 (56.8)	62/83 (74.7)	58/90 (64.4)
ESBL + Enterobacteriaceae	48/84 (57.1)	45/73 (61.6)	33/45 (73.3)	27/39 (69.2)
<i>Enterobacter cloacae</i>	10/17 (58.8)	4/16 (25.0)	4/7 (57.1)	3/8 (37.5)
<i>Escherichia coli</i>	32/51 (62.7)	26/42 (61.9)	17/23 (73.9)	16/23 (69.6)
ESBL + <i>Escherichia coli</i>	11/20 (55.0)	5/10 (50.0)	8/12 (66.7)	5/7 (71.4)
<i>Klebsiella oxytoca</i>	9/14 (64.3)	7/12 (58.3)	7/8 (87.5)	4/7 (57.1)
<i>Klebsiella pneumoniae</i>	53/86 (61.6)	58/91 (63.7)	32/42 (76.2)	33/48 (68.8)
ESBL + <i>Klebsiella pneumoniae</i>	31/53 (58.5)	34/52 (65.4)	22/30 (73.3)	19/27 (70.4)
<i>Proteus mirabilis</i>	13/24 (54.2)	11/20 (55.0)	9/11 (81.8)	7/10 (70.0)
ESBL + <i>Proteus mirabilis</i>	5/10 (50.0)	7/11 (63.6)	4/5 (80.0)	5/6 (83.3)
<i>Serratia marcescens</i>	9/18 (50.0)	7/12 (58.3)	4/5 (80.0)	3/6 (50.0)
<i>Haemophilus influenzae</i>	19/22 (86.4)	8/16 (50.0)	11/12 (91.7)	4/8 (50.0)

Table 17: Microbiologic Response Rates by Baseline Pathogen from a Phase 3 Study of Nosocomial Pneumonia (mITT and ME populations)

Baseline Pathogen Category Baseline Pathogen	mITT Population		ME Population	
	ZERBAXA n/N (%)	Meropenem n/N (%)	ZERBAXA n/N (%)	Meropenem n/N (%)

<i>Pseudomonas aeruginosa</i>	47/63 (74.6)	41/65 (63.1)	23/29 (79.3)	21/38 (55.3)
AmpC Overexpressing <i>Pseudomonas aeruginosa</i>	6/9 (66.7)	1/6 (16.7)	2/4 (50.0)	1/5 (20.0)
Enterobacteriaceae	145/195 (74.4)	129/185 (69.7)	57/83 (68.7)	59/90 (65.6)
ESBL + Enterobacteriaceae	56/84 (66.7)	52/73 (71.2)	30/45 (66.7)	27/39 (69.2)
<i>Enterobacter cloacae</i>	11/17 (64.7)	8/16 (50.0)	4/7 (57.1)	6/8 (75.0)
<i>Escherichia coli</i>	43/51 (84.3)	33/42 (78.6)	18/23 (78.3)	17/23 (73.9)
ESBL + <i>Escherichia coli</i>	18/20 (90.0)	8/10 (80.0)	10/12 (83.3)	6/7 (85.7)
<i>Klebsiella (Enterobacter)</i> <i>aerogenes</i>	6/8 (75.0)	6/8 (75.0)	1/1 (100)	1/1 (100)
<i>Klebsiella oxytoca</i>	13/14 (92.9)	8/12 (66.7)	7/8 (87.5)	4/7 (57.1)
<i>Klebsiella pneumoniae</i>	63/86 (73.3)	65/91 (71.4)	30/42 (71.4)	32/48 (66.7)
ESBL + <i>Klebsiella</i> <i>pneumoniae</i>	33/53 (62.3)	38/52 (73.1)	20/30 (66.7)	18/27 (66.7)
<i>Proteus mirabilis</i>	18/24 (75.0)	14/20 (70.0)	7/11 (63.6)	7/10 (70.0)
ESBL + <i>Proteus mirabilis</i>	7/10 (70.0)	7/11 (63.6)	3/5 (60.0)	5/6 (83.3)
<i>Serratia marcescens</i>	11/18 (61.1)	9/12 (75.0)	2/5 (40.0)	3/6 (50.0)
<i>Haemophilus influenzae</i>	20/22 (90.9)	11/16 (68.8)	11/12 (91.7)	4/8 (50.0)

In the mITT population, per subject microbiologic cure was achieved in 193/264 (73.1%) of ZERBAXA-treated patients and in 168/247 (68.0%) of meropenem-treated patients. Similar results were achieved in the ME population in 81/115 (70.4%) and 74/118 (62.7%) patients, respectively.

In a subset of Enterobacteriaceae isolates from both arms of the trial that met pre-specified criteria for beta-lactam susceptibility, genotypic testing identified certain ESBL groups (e.g., TEM, SHV, CTX-M, OXA) in 157/511 (30.7%). Cure rates in this subset were similar to the overall trial results.

13 REFERENCES

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14 HOW SUPPLIED/STORAGE AND HANDLING

14.1 How Supplied

ZERBAXA 1.5 g (ceftolozane and tazobactam) for injection is supplied in single-dose vials containing ceftolozane 1 g (equivalent to 1.147 g of ceftolozane sulfate) and tazobactam 0.5 g (equivalent to 0.537 g of tazobactam sodium) per vial. Vials are supplied in cartons containing 10 vials.

14.2 Storage and Handling

ZERBAXA vials should be stored refrigerated at 2 to 8°C (36 to 46°F) and protected from light.

The reconstituted solution, once diluted, may be stored for 24 hours at room temperature or for 7 days under refrigeration at 2 to 8° C (36 to 46°F).

15 PATIENT COUNSELING INFORMATION

Serious Allergic Reactions

Advise patient that allergic reactions, including serious allergic reactions, could occur and that serious reactions require immediate treatment. Ask patient about any previous hypersensitivity reactions to ZERBAXA, other beta-lactams (including cephalosporins) or other allergens [*see Warnings and Precautions (5.2)*].

Potentially Serious Diarrhea

Advise patient that diarrhea is a common problem caused by antibacterial drugs. Sometimes, frequent watery or bloody diarrhea may occur and may be a sign of a more serious intestinal infection. If severe watery or bloody diarrhea develops, tell patient to contact his or her healthcare provider *[see Warnings and Precautions (5.3)]*.

Antibacterial Resistance

Counsel patient that antibacterial drugs including ZERBAXA should only be used to treat bacterial infections. They do not treat viral infections (e.g., the common cold). When ZERBAXA is prescribed to treat a bacterial infection, patients should be told that although it is common to feel better early in the course of therapy, the medication should be taken exactly as directed. Skipping doses or not completing the full course of therapy may (1) decrease the effectiveness of the immediate treatment and (2) increase the likelihood that bacteria will develop resistance and will not be treatable by ZERBAXA or other antibacterial drugs in the future *[see Warnings and Precautions (5.4)]*.

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